

# Determination of Chlorite— Quick 2-step method

DOC316.52.93149

Based on Standard Methods 4500-ClO<sub>2</sub> E  
for drinking water and wastewater

This application note covers the following applications:

| Method                                  | Range   | Titrant      | Buffer, KI and Acid              | Sample volume |
|---|---|--------------|----------------------------------|---------------|
| <b>Chlorite (H)<br/>(Quick, 2-step)</b> | 0.100 to 5.00 mg ClO <sub>2</sub> <sup>-</sup> /L<br>Cl <sub>2</sub> more than 0.100 mg/L | 0.00564N PAO | 1 mL pH 7<br>1g<br>2 mL 2.5N HCl | 200 mL        |
| <b>Chlorite (L)<br/>(Quick, 2-step)</b> | 0.100 to 5.00 mg ClO <sub>2</sub> <sup>-</sup> /L<br>Cl <sub>2</sub> less than 0.100 mg/L | 0.00564N PAO | 1 mL pH 7<br>1g<br>2 mL 2.5N HCl | 200 mL        |

## 1. Important information

- The AT1000 is factory programmed to use the 10-mL syringe. The method uses a 5-mL syringe. Before analysis, make sure to change the syringe volume on the instrument. Refer to [10.1 Changing the syringe volume on the AT1000](#).
- Treat glassware to decrease chlorine demand before analysis. Soak glassware in dilute bleach solution for a minimum of 1 hour. Rinse thoroughly with deionized water. Use the glassware for this method only.
- Minimize agitation when measuring sample volumes. Remove sample portions with a volumetric pipette. Always put the tip of the pipette at the bottom of the sample container. If using 200 mL sample portions, use a 100-mL volumetric pipette to withdraw two portions of sample.
- Always use organic free water for sample dilution.
- Rinse the electrode and anti-diffusion tip with DI water before every titration.
- Purge the syringe each day before the analysis.
- Do not substitute buffers designed for calibrating pH meters. They contain dyes that can interfere with amperometric titration.
- Do not use buffers contaminated with mold or bacteria.
- Clean the electrode periodically Refer to [10.2 Cleaning the Electrode](#). Clean and correctly maintained electrodes are necessary to get sharp amperometric endpoints. Clean the electrode when noise in the titration curve interferes with detection of the endpoint. The electrode cleaning duration is approximately 10 minutes. Always clean new electrodes before the analysis.
- The electrode orientation is very important. Noise that occurs when the electrode is not correctly oriented can interfere with accurate detection of the equivalence point. Refer to [4.1 Position of the electrode and injection tips](#) for information on electrode and delivery tip positioning for this method.
- A fast stirring can pull air into the sample and bubbles may get caught on the electrode tip. Air bubbles on the electrode tip have a negative effect on the analysis results. Adjust the stirring speed during a titration with the up and down arrows on the instrument. Alternatively, change the stirring speed in the method edit window.
- The method is programmed to measure **200 mL** of sample. **To use less than 200 mL of sample, change the sample volume at the method edit window.** Refer to [10 Appendix](#) for more information.

## 2. Introduction

This application is based on the Standard Methods 4500-ClO<sub>2</sub> E, an amperometric method which distinguishes three different compounds: chlorine dioxide (ClO<sub>2</sub>), free chlorine (Cl<sub>2</sub>) and chlorite (ClO<sub>2</sub><sup>-</sup>).

**This method describes a quick 2-step method for chlorite only.** Both the L and H methods **cover the same range in chlorite.** The difference in the methods is in the range for Titration 1. The methods are as follows:

|                                   |  |  |
|-----------------------------------|--|--|
| ClO <sub>2</sub> <sup>-</sup> (H) | 0.100 – 5.00 mg ClO <sub>2</sub> <sup>-</sup> /L | Cl <sub>2</sub> <b>above</b> 0.100 mg (titration 1)    |
| ClO <sub>2</sub> <sup>-</sup> (L) | 0.100 – 5.00 mg ClO <sub>2</sub> <sup>-</sup> /L | Cl <sub>2</sub> <b>less than</b> 0.100mg (titration 1) |

### 3. Principle

Two amperometric titrations are done on one titration sample. The results of each titration are stored and at the end of the sequence the concentration of chlorite ( $\text{ClO}_2^-$ ) is displayed.

The sample pH is adjusted to 7 by addition of buffer, and the sample is then degassed with nitrogen to remove chlorine that might be present. An excess of KI is added and the titration is launched to neutralize any  $\text{Cl}_2$  not volatilized by the degassing. Next, concentrated hydrochloric acid is added to the cell to give the  $\text{ClO}_2^-$  determined during the second titration.

The table below gives details of the sequence:

|             |   |
|-------------|---|
| Titration 1 | $\text{Cl}_2$ not volatilized by the nitrogen gas purge |
| Titration 2 | $\text{ClO}_2^-$  |

### 4. Electrode and reagents

**Electrode:** Pt-Pt electrode with temperature sensor, IntelliCAL MTC695

**Titrant:** Phenyl Arsine Oxide (PAO) 0.00564 eq/L solution

**Reagents:** pH 7 phosphate buffer  
Potassium iodide (KI) powder  
Hydrochloric acid (HCl) 2.5 N solution

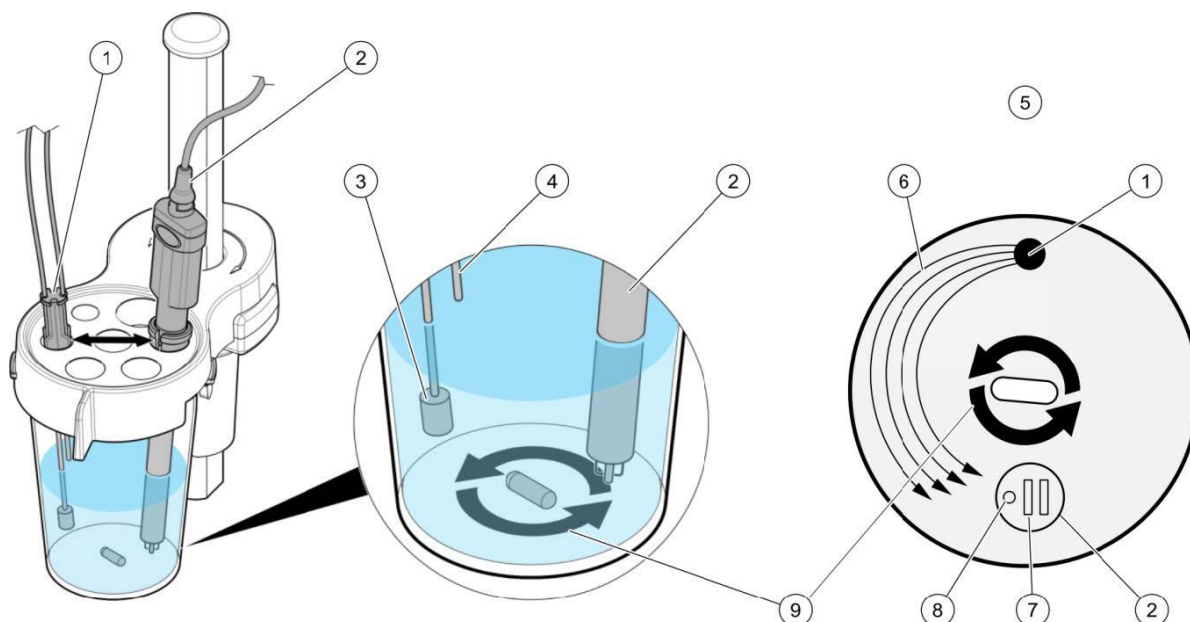
**Deionized water**

#### 4.1. Position of the electrode and injection tips

The position of the electrode and injection tips in the titration cell is very important in this application. **If the electrode is incorrectly positioned, noise in the titration curve can adversely affect the results.**

Refer to the steps and the figure that follows to correctly position the electrode and injection tips.

1. Put the electrode in the opposite hole of the tubes in the sensor holder (items 1 and 2 in figure).
2. Turn the electrode so that the platinum wires are perpendicular to the sample flow and the temperature sensor is before the platinum wires (items 6 to 8 in figure).
3. Put the tube from the pump above the sample surface (item 4 in figure).
4. Make sure that the tube with the anti-diffusion tip is fully into the sample (item 3 in figure).



|                       |                       |                       |
|-----------------------|-----------------------|-----------------------|
| 1. Tube holder        | 4. Tube from the pump | 7. Platinum wires     |
| 2. Electrode          | 5. Top view           | 8. Temperature sensor |
| 3. Anti-diffusion tip | 6. Flow direction     | 9. Stirring direction |

## 5. Settings

### 5.1. Chlorite determination

Two methods, Chlorine Dioxide L and Chlorine Dioxide H are available. The difference between **L** and **H** are the settings for **Titration 1**.

If the expected equivalent volume for titration 1 is very low (< 0.1 mL), use the **L** applications. If not, use the **H** applications. The ranges for Chlorine Dioxide, Chlorite, and Chlorine are the same for the two methods.

The settings below have been defined with:

- Sample volume: 200 mL
- Titrant concentration: 0.00564 eq/L Phenyl Arsine Oxide
- Continuous imposed voltage: 100 mV (reversed at each analysis)
- Syringe volume: **5 mL**  
The default syringe volume for the AT1000 is set to 10 mL. These applications need a 5-mL syringe. When loading an application, if the message syringe to replace is displayed, change the syringe volume in the Syringe management option of the Maintenance menu. Refer to [10 Appendix](#) for more information.
- If a blank is tested, analyze this as a sample. The BLANK option is not compatible with this method.

| Name                                      | Default parameter  |            | Unit    |
|---|--|------------|---------|
| Application name                          | Chlorite L   | Chlorite H |         |
| <b>Syringe</b>                            |  |            |         |
| Advisable                                 | 5 mL (Hamilton)  |            |         |
| <b>Sample</b>                             |  |            |         |
| Name                                      | Water ? <sup>1</sup>   |            |         |
| Amount                                    | 200  |            | mL      |
| <b>QC</b>                                 |  |            |         |
| Name                                      | QC Sample  |            |         |
| <b>Probe</b>                              |  |            |         |
| Recommended electrode                     | MTC695   |            |         |
| <b>Titrant PAO 0.00564 N</b>              |  |            |         |
| Real concentration                        | 0.00564  |            | eq/L    |
| <b>Sample preparation 1 (titration 1)</b> |  |            |         |
| Active                                    | Yes  |            |         |
| Message                                   | Sample amount: 200 mL in GWB and press OK                                    |            |         |
| Stirring speed                            | 0  |            | %       |
| <b>Sample preparation 2 (titration 1)</b> |  |            |         |
| Active                                    | Yes  |            |         |
| Message                                   | Add 1.0 mL of buffer pH 7 then start purge with N <sub>2</sub> and press OK  |            |         |
| Stirring speed                            | 0  |            | %       |
| <b>Purge (titration 1)</b>                |  |            |         |
| Active                                    | Yes  |            |         |
| Time                                      | 15   |            | minutes |
| Stirring speed                            | 0  |            | %       |
| Message                                   | Purge in progress. Please wait...  |            |         |
| <b>Sample preparation 3 (titration 1)</b> |  |            |         |
| Active                                    | Yes  |            |         |
| Message                                   | Pour the purged sample into a titration beaker – Add a stir bar and press OK |            |         |
| Stirring speed                            | 0  |            | %       |
| <b>Manual addition 1 (titration 1)</b>    |  |            |         |
| Active                                    | Yes  |            |         |
| Message                                   | Place the sample on the instrument - Add <b>1.0 g</b> of KI and press OK     |            |         |
| Stirring speed                            | 0  |            | %       |
| <b>Reagents mixing (titration 1)</b>      |  |            |         |
| Active                                    | Yes  |            |         |
| Time                                      | 5  |            | seconds |
| Stirring speed                            | 1  |            | %       |
| Message                                   | Reagents mixing. Please wait...  |            |         |
| <b>Dip electrode (titration 1)</b>        |  |            |         |

<sup>1</sup> ? in the name, shows that the sample name will be automatically incremented with a number for each analysis

| Name                                   | Default parameter  |            | Unit    |
|--|--|------------|---------|
|  | Chlorite L   | Chlorite H |         |
| Application name                       |  |            |         |
| Active                                 | Yes  |            |         |
| Message                                | Dip electrode in sample and press OK   |            |         |
| Stirring speed                         | 1  |            | %       |
| <b>Titration 1</b>                     |  |            |         |
| Active                                 | Yes  |            |         |
| Stirring speed                         | 1  |            | %       |
| Predose ordinate                       | 0.1  | 0.4        | µA      |
| Predose speed                          | 0.3  | 2.0        | mL/min  |
| Delay                                  | 20   |            | seconds |
| Max. vol. stop point                   | 5  |            | mL      |
| Stop on last EQP                       | Yes  |            |         |
| Increment size                         | 0.001  | 0.010      | mL      |
| EQP min. ordinate                      | -0.03  | -0.1       | µA      |
| EQP max. ordinate                      | 0.03   | 0.2        | µA      |
| Result 1 name                          | Intermediate 3   |            | mL      |
| R1 hide                                | Yes  |            |         |
| R1 min                                 | 0  |            | mL      |
| R1 max                                 | 5  |            | mL      |
| R1 QC min                              | 0  |            | mL      |
| R1 QC max                              | 5  |            | mL      |
| Result 2 name                          | C  |            |         |
| R2 hide                                | Yes  |            |         |
| R2min                                  | 0  |            | mL/mL   |
| R2max                                  | 0.025  |            | mL/mL   |
| R2QC min                               | 0  |            | mL/mL   |
| R2 QC max                              | 0.025  |            | mL/mL   |
| R2 equation                            | FX*(R1/SA) = G3  |            |         |
| R2 user value                          | 1  |            |         |
| <b>Manual addition 2 (titration 2)</b> |  |            |         |
| Active                                 | Yes  |            |         |
| Message                                | Add 2.0 mL of HCl 2.5 N then place the solution in the dark and press OK         |            |         |
| Stirring speed                         | 1  |            | %       |
| <b>Reaction (titration 2)</b>          |  |            |         |
| Active                                 | Yes  |            |         |
| Message                                | 5  |            | minutes |
| Stirring speed                         | 0  |            | %       |
| Message                                | Dark reaction in progress. Please wait...  |            |         |
| <b>Dip electrode (titration 2)</b>     |  |            |         |
| Active                                 | Yes  |            |         |
| Message                                | Place the sample on the instrument then dip the electrode in sample and press OK |            |         |
| Stirring speed                         | 0  |            | %       |
| <b>Titration 2</b>                     |  |            |         |
| Active                                 | Yes  |            |         |
| Stirring speed                         | 1  |            | %       |
| Predose ordinate                       | 1.0  |            | µA      |
| Predose speed                          | 2.5  |            | mL/min  |
| Delay                                  | 20   |            | seconds |
| Max. vol. stop point                   | 15   |            | mL      |
| Stop on last EQP                       | Yes  |            |         |
| Increment size                         | 0.010  |            | mL      |
| EQP min. ordinate                      | -0.1   |            | µA      |
| EQP max. ordinate                      | 0.2  |            | µA      |
| Result 3 name                          | Intermediate 4   |            | mL]     |
| R3 min                                 | 0  |            | mL/mL   |
| R3 max                                 | 0.075  |            | mL/mL   |
| R3QC min                               | 0  |            | mL/mL   |
| R3QC max                               | 0.075  |            | mL/mL   |
| R3 equation                            | FX*(R1/SA) = G4  |            |         |
| R3 user value                          | 1  |            |         |
| Result 3 name                          | D  |            |         |

| Name             | Default parameter                 |            | Unit                   |
|------------------|-----------------------------------|------------|------------------------|
|                  | Chlorite L                        | Chlorite H |                        |
| Application name |                                   |            |                        |
| Result 3 hide    | Yes                               |            |                        |
| Result 4 name    | Chlorite                          |            |                        |
| R4 hide          | No                                |            |                        |
| R4 min           | 0                                 |            | mg ClO <sub>2</sub> /L |
| R4 max           | 6                                 |            | mg ClO <sub>2</sub> /L |
| R4QC min         | 0                                 |            | mg ClO <sub>2</sub> /L |
| R4 QC max        | 6                                 |            | mg ClO <sub>2</sub> /L |
| R4 equation      | V1/V1*FX*(G4+(F3/SA)-G3)*TC*16863 |            |                        |
| R4 user value    | 1                                 |            |                        |

## 5.2. Recommendations for modification of the settings

Some parameters can be adjusted, but this is mainly for analysis time reduction. It should be noted that the impact of any adjustments can be a loss of precision on the results.

### 5.2.1. Sample preparation messages

Messages for sample preparation can be removed from the sequence by setting **No** in the field **Active** in the message section. In this way, the instrument will not give information about sample preparation during the analysis sequence. Methods which can be deactivated are the following:

- Automatic addition 1 (titration 1)
- Manual addition 1 (titration 1)
- Manual addition 2 (titration 1)
- Manual addition 3 (titration 2)
- Sample preparation 1 (titration 1)
- Sample preparation 2 (titration 1)
- Purge (titration 1)
- Sample preparation 3 (titration 1)
- Manual addition 1 (titration 1)
- Manual addition 2 (titration 2)
- Reaction (titration 2)

**Note:** It is not recommended to change the increment sizes because they have been optimized for the best equivalent point detection.

### 5.2.2. Pre-doses in ordinate

Predoses in ordinate are used to decrease the titration duration. They have been fixed for all titrations for chlorine dioxide and chlorite applications. Their parameters (**Predose ordinate** and **Predose speed**) have been set empirically and are system dependent. A titration starting with an ordinate under the target can happen but does not have an impact on the result. The table that follow shows some indications.

| Observation  | Resolution  |
|--|---|
| The titration is still too long (too many points before inflection).   | Decrease the predose ordinate in the ordinate section or increase the titrant addition speed (no more than 2.5 mL/min). |
| The initial point of the titration curve is too low (not enough points before inflection) and the EQP is not detected. | Decrease the titrant addition speed in the ordinate section or increase the predose ordinate.                           |

## 6. Procedure

### 6.1. Before starting

**NOTICE: All glassware must be treated for chlorine demand before any analysis.**

- Soak all glassware in dilute bleach solution for at least 1 hour. Rinse thoroughly with deionized water. Use the glassware for this method only.
- Minimize agitation when measuring sample volumes. Remove sample portions with a volumetric pipette. Always put the tip at the bottom of the sample container. If using 200 mL sample increments, use a 100 mL pipette to withdraw two portions of sample.
- Always use organic free water for sample dilution.
- Do not substitute buffers designed for calibrating pH meters. They may contain dyes that interfere in amperometric titration
- Do not use buffers contaminated with mold or bacteria.

- Rinse the electrode and anti-diffusion tip with deionized water before every titration.

## 6.2. Sample analysis

1. Use a pipette to add 200 mL of sample into a Gas Washing Bottle (GWB).
2. Add 1.0 mL of phosphate buffer pH 7 and swirl to mix.
3. Insert the purge tube and dispersion tip into the GWB. Connect the GWB inlet to a tank of purified nitrogen.
4. Use a needle valve to adjust the flow of nitrogen to provide a steady stream of bubbles through the sample.
5. Purge the nitrogen gas through the sample for 15 minutes.
6. After the purge, transfer the purged sample to a 250-mL glass beaker. Add a magnetic stir bar and put in the instrument.
7. Add 1.0 g of KI.
8. The reagents are mixed. Dip the electrode and addition tip into the sample.
9. Titration 1 starts.
10. When titration 1 is finished, raise the electrode holder.
11. Add 2.0 mL of 2.5 N hydrochloric acid (HCl) and stir for a few seconds.
12. Carefully remove the sample from the stirrer and put the sample in a dark environment.
13. Wait 5 minutes for the reaction.
14. At the end of the 5 minutes, carefully put the sample back onto the instrument.
15. Dip the electrode and addition tip into the sample.
16. Titration 2 starts.
17. At the end of titration 2, the results are shown.

## 7. Results

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### 7.1. Displayed Results

At the end of the analysis sequence the following result is displayed:

$\text{ClO}_2^-$  in mg/L as  $\text{ClO}_2^-$

### 7.2. Results calculation

$\text{ClO}_2^-$  calculation:

$$\text{ClO}_2^- = D \times N \times 16863$$

Where:

D = Result of titration 2 (mL titrant at equivalent point/mL of sample)

N = Concentration of the titrant

## 8. Examples of chlorite determination

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The results described below are indicative and obtained for a given sample in optimized conditions following good laboratory practices. These indicative values are sample-dependent, electrode-dependent and operating cell-dependent.

Results for four determinations of a synthetic mixture:

**Sample:** 200 mL of solution

**Application:** Chlorite L

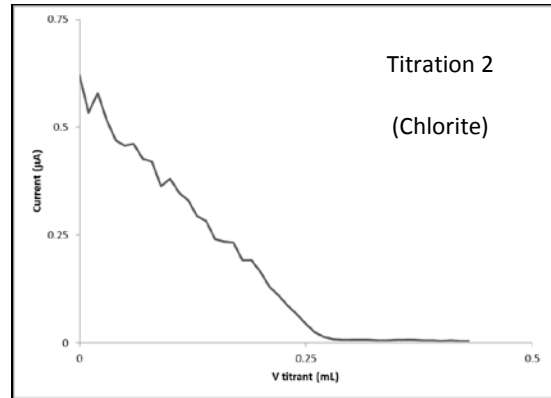
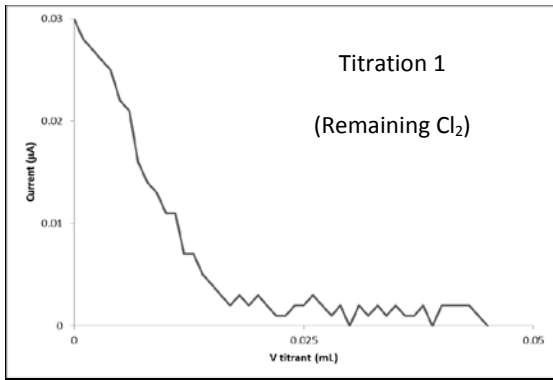
**Temperature of analysis:** Room temperature

**Mean values:**  $\text{ClO}_2^-$ : 0.154 mg/L  
 $\text{Cl}_2$ : -

**Standard deviation:**  $\text{ClO}_2^-$ : 0.008 mg/L  
 $\text{Cl}_2$ : -

**Relative standard deviation:**  $\text{ClO}_2^-$ : 5.21%  
 $\text{Cl}_2$ : -

**Example of titration curves:**



## 9. Bibliography

- *Standard Methods 4500-ClO<sub>2</sub> E*
- *AutoCAT 9000 Manual 50081 3rd edition*

## 10. Appendix

### 10.1. Changing the syringe volume on the AT1000.

The AT1000 instrument is delivered with the syringe volume set to 10 mL. The amperometric applications require a 5-mL syringe volume. The syringe volume must be changed before the applications can be started. Complete the steps that follow to change the syringe volume:

1. From the HOME screen select MAINTENANCE > SYRINGE MANAGEMENT > SYRINGE VOLUME CHANGE.

**Note:** If the AT1000 instrument has 2 syringes, select the syringe to edit.

2. Use the arrow keys to select 5 ML (HAMILTON), then push **SELECT**. The display shows APPLYING 5ML (HAMILTON) SETTINGS followed by SYRINGE VOLUME UPDATED.
3. Push **OK**.
4. Push **HOME** to go back to the HOME screen.

### 10.2. Cleaning the Electrode

This procedure should be done before first use, after dry storage, and when the electrode response is slowed or equivalence points are missed.

1. Prepare a cleaning solution of 20-mL HNO<sub>3</sub>/100 mL. Always add acid to water! Always wear personal protective equipment!
  2. From the HOME screen, select MAINTENANCE > CLEAN PT-PT ELECTRODE
  3. Pour enough solution in the beaker to cover the electrode.
  4. Select OK
- Note:** If the stirrer does not start, push the up and down arrows.
5. After five minutes, when prompted, rinse the electrode with DI water and fill the beaker with enough tap water to cover the electrode
  6. Put the PtPt electrode in the water and select OK.
  7. After five minutes, the cleaning is complete.

### 10.3. Hide or Show a Result

1. From the Home screen, select SETTINGS>APPLICATIONS.
2. Select EDIT from the list of actions.
3. Highlight the application and push **EDIT**.
4. Use the down arrow to go to METHOD>RESULTS.
5. Select YES or NO to show or hide results:
  - **Yes**—the result is not shown (hidden) at the end of the titration
  - **No**—the result is shown at the end of the titration

### 10.4. Titrant calibration

#### 10.4.1. Set the Titer directly from the C.O.A.

Before the Titer is entered directly from the certificate of analysis (C.O.A.) refer to the customer laboratory standard operation procedure (S.O.P.) to determine if this is acceptable.

1. From the HOME screen, select SETTINGS>APPLICATIONS>EDIT

2. Select the application for the titer
3. Scroll down in the application to Titrant
4. Select REAL CONCENTRATION
5. Using the arrow keys or a keyboard, enter the titer value from the C.O.A.
6. Select OK
7. Go back to the Home screen.

**Note:** This step is needed only once for each syringe, even if there is more than one method associated with it. Enter the value from the C.O.A. using the up and down arrow keys or a USB Keyboard. Go back to the Home Screen.

#### 10.4.2. Calibration of the titrant with 0.0282N Iodine

The 0.000564N PAO titrant is calibrated against a standard solution of 0.0282 N Iodine



The iodine solution can also be calibrated. The procedure is described in the in the Sulfite working procedures, which are included on the Amperometric titration method key.

If the standard iodine concentration given in the Certificate of Analysis (or obtained by calibration) is different from the default concentration of 0.0282 N, the real value must be manually entered as the concentration of the standard.

##### 10.4.2.1. Procedure

Accurately pipette 0.5 mL of iodine standard solution 0.0282 N and dilute it to 200 mL with deionized water.

Calibrate the titrant using the titrant calibration option instead of the sample analysis. Add KI powder and pH 4 when required. On a titrator with 2 pumps, pH 4 buffer is pumped using Pump 2.

##### 10.4.2.2. Results

The results described below are indicative and obtained following good laboratory practices. These indicative values are sample-dependent, electrode-dependent and operating cell-dependent.

The instrument calculates the titrant concentration directly in eq/L.

$$C_{(PAO)} = \frac{V_{(I2)} * C_{(I2)}}{V_{(PAO)}}$$

- C<sub>(PAO)</sub>: Concentration of titrant: Phenylarsine Oxide (PAO) in eq/L,
- C<sub>(I2)</sub>: Concentration of standard: Iodine (I2) in eq/L, currently 0.0282 eq/L
- V<sub>(I2)</sub>: Volume of standard: Iodine (I2) in mL, currently 0.5 mL
- V<sub>(PAO)</sub>: Volume of the titrant: Phenylarsine Oxide (PAO) in mL added to reach the equivalent point

Experimental conditions:

- **Burette volume:** 5 mL
- **Sample:** 200 mL of deionized water with 0.5 mL of standard solution iodine 0.0282 eq/L
- **Addition:** 0.1 g KI and 1 mL buffer pH 4
- **Titrant:** PAO 0.000564 eq/L
- **Acceptable Range:** 0.000564N +/- 10% (0.000508-0.000620N)

Settings:

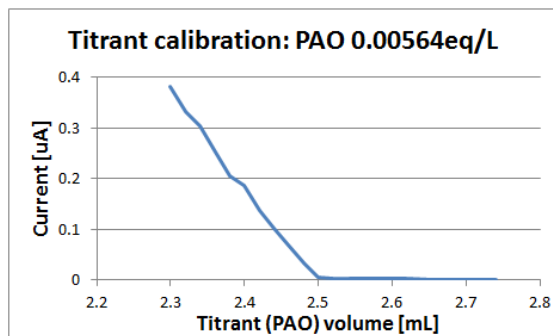
Results for 5 replicates of the titrant:

|                              |         |      |
|------------------------------|---------|------|
| <b>Average concentration</b> | 0.00561 | eq/L |
| <b>SD</b>                    | 0.00002 | eq/L |
| <b>RSD</b>                   | 0.4     | %    |

Titration curve: µA vs. volume of titrant



- **Settings:** Refer to [10.4.2.3 Titrant calibration settings \(default parameters\)](#)
- **Number of determinations:** 5 samples
- **Temperature of analysis:** Room temperature



#### 10.4.2.3. Titrant calibration settings (default parameters)

|                                   | Setting | Unit    |
|-----------------------------------|---------|---------|
| Titrant name                      | PAO     |         |
| Nominal concentration             | 0.00564 | eq/L    |
| Calibration frequency             | 0       | days    |
| Stirring speed (%)                | 1       | %       |
| Predose volume                    | 2.1     | mL      |
| Delay                             | 20      | seconds |
| Stop on last EQP                  | Yes     |         |
| Min increment size                | 0.02    | mL      |
| Max increment size                | 0.05    | mL      |
| EQP min. ordinate                 | -0.1    | µA      |
| EQP max. ordinate                 | 0.2     | µA      |
| <b>Titrant calibration result</b> |         |         |
| Min. titrant concentration        | 0.0055  | eq/L    |
| Max. titrant concentration        | 0.0058  | eq/L    |
| <b>Standard</b>                   |         |         |
| Name                              | Iodine  |         |
| Amount                            | 0.500   | mL      |
| Min amount                        | 0.490   | mL      |
| Max amount                        | 0.510   | mL      |
| Concentration                     | 0.0282  | eq/L    |

#### 10.4.2.4. Modification of the parameters for the titrant calibration

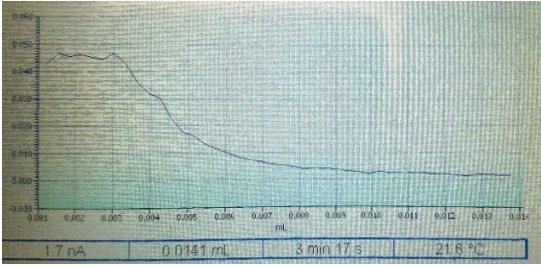



The titrant calibration application has been optimized for an amount of standard higher than 0.49mL, a standard concentration higher than 0.0270 eq/L and a titrant concentration between 0.0055 eq/L and 0.0058 eq/L.

Based on the concentration of the standard, the titrant volume needed for the equivalence will be affected by an amount or a concentration of the standard different to the default values. The predose volume can be adjusted in relation to this amount, to ensure about 0.2 mL of titrant before the equivalence point.

As an example, the table below shows the effect of the standard concentration on the equivalent volume and the optimum predose volume as a function of the equivalent volume expected.

| Standard volume and concentration | Titrant concentration | Theoretical equivalent titrant volume | Number of addition points before equivalent point detection with default predose at 2.1mL | Optimized predose volume |
|-----------------------------------|-----------------------|---------------------------------------|---|--------------------------|
| 0.50 mL at 0.0270 eq/L            | 0.0058 eq/L           | 2.33 mL                               | 11  | 2.1 mL                   |
| 0.50 mL at 0.0270 eq/L            | 0.0055 eq/L           | 2.45 mL                               | 18  | 2.2 mL                   |
| 0.50 mL at 0.0290 eq/L            | 0.0058 eq/L           | 2.50 mL                               | 20  | 2.3 mL                   |
| 0.50 mL at 0.0290 eq/L            | 0.0055 eq/L           | 2.64 mL                               | 27  | 2.4 mL                   |

## 11. Troubleshooting

| Symptom   | Probable cause   | Solution  |
|---|--|---|
| <p>No clear equivalence point, equivalence point not found.</p>   |    | <p>Concentration too low?<br/>Do a cleaning procedure<br/>After cleaning, analyze a mid-range standard to verify performance.</p>   |
| <p>Titration curve is noisy.<br/>No or incorrect equivalence point found.<br/>Electrode responds slowly; titration takes longer than usual.</p> |    | <p>Concentration too low?<br/>Check for bubble caught on electrode<br/>Verify electrode is properly oriented<br/>Clean electrode</p>  |
| <p>Predose exceeds the set ordinate.<br/>Electrode responds slowly; titration takes longer than usual.</p>                                      | <p>Dirty or polarized electrode</p>  | <p>The analyte level is too low to be detected.<br/>Clean the electrode.</p>  |
| <p>There are bubbles on the electrode tip.</p>  |  | <p>Picture on the left shows no bubbles. Picture on the right shows a bubble caught on the electrode.<br/>Adjust the stirring speed to 35-40% which will not normally cause bubbles to occur.<br/>If analyzing a standard, make sure that the volume of water used is sufficient.</p> |
| <p>Flat signal<br/>Noisy signal<br/>Electrode is dirty.</p>   |  | <p>No analyte (blank)<br/>No buffer added<br/>No KI added<br/>Clean the electrode.</p>  |

## 11.1. Waste management

The laboratory has the responsibility to follow all of the federal, state, and local regulations governing waste management (particularly the hazardous waste identification rules) and land disposal restrictions. The laboratory must minimize and control all releases from fume hoods and bench operation to protect the air, water and land. Compliance with all sewage discharge permits and regulations is also required.

For more information on waste management refer to the *Waste Management Manual for Laboratory Personnel* guide, available from the American Chemical Society's Department of Government Regulations and Science Policy, 1155 16th Street N. W., Washington D. C. 20036, (202) 872-4477.

## 12. Parts List

| Description   | Quantity per test | Unit   | Item no. |
|---|-------------------|--------|----------|
| <b>Required reagents</b>  |                   |        |          |
| Phenylarsine oxide (PAO) titrant, 0.00564 N                                 | varies            | 1 L    | 199953   |
| Buffer solution, pH 7 (automatic addition)                                  | About 2 mL        | 1 L    | 2155353  |
| Acetate buffer solution, pH 7 with dropper (manual addition)                | About 2 mL        | 100 mL | 2155332  |
| Potassium iodide, ACS or better <sup>2</sup>                                | 0.1 g             | 100 g  | 16726H   |
| Swiftest™ dispenser, with refill vial                                       | varies            | each   | 2834100  |
| Refill vial, Siwftest dispenser   | varies            | 0.1 g  | 2105660  |
| Nitric acid, 1 :1   | 20 mL             | 500 mL | 254049   |
| <b>Required equipment</b>   |                   |        |          |
| 0.1-g scoop for addition of KI to the sample                                | 1                 | each   | 2657201  |
| Beaker, low form Griffin, glass, 250 mL,                                    | 1                 | each   | 50046H   |
| Beaker, low form Griffin, glass, 250 mL                                     | 1                 | 12/pkg | 50076H   |
| Cylinder, graduated, 250 mL   | 1                 | each   | 50846    |
| Magnetic stir bar, PTFE coated, 2 x 3/8 in.                                 | 1                 | each   | 5008500  |
| Gas washing bottle, 1200 mL   | 1                 | each   | 2662200  |
| <b>Optional reagents</b>  |                   |        |          |
| Chlorine Standard Solution, Voluette® ampules, 50–75 mg/L 16, 10 mL ampules | varies            | 16/pkg | 1426810  |
| Chlorine standard solution, Pour Rite® ampules, 25–30 mg/L 20, 2 mL ampules | varies            | 20/pkg | 2630020  |
| Dilution Water, ASTM Type III organic-free                                  | varies            | 500 mL | 2641549  |

<sup>2</sup> The KI granules must be ground with a mortar and pestle for this method if the Swiftest is not used.

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