Determination of Ammonia

Introduction

This application describes the distillation of samples that contain ammonia and the subsequent determination using the ammonia cuvette test TNT830, 831 or 832. The procedure follows USEPA methods 350.1, 351.1, 351.2.

Material:

• MDD002 - Micro Dist Reactor Block

Or

- DRB20004 DRB200 Reactor Block (8 x 20-mm wells)
 - LZT144 DRB200 Adapter Sleeve for Micro Dist (8/pkg)
- A17517A Micro Dist Ammonia tubes (user filled, 50/pkg)
- A17117A Micro Dist Ammonia tubes (user filled, 100/pkg)
- 17023L Press
- 17013L Protective gloves, large
- 17013S Protective gloves, small
- 17012 Stands, collection tubes
- 21302 Stands, sample tubes
- BBP065 Pipet, adjustable volume, 1.0 5.0 mL
- BBP068 Pipet tips, for 1.0 5.0 mL pipet
- 1970010 Pipet, TenSette®, 1.0 10.0 mL
- 2199796 Pipet tips for TenSette®, 1.0 10.0 mL
- TNT830 ULR Ammonia TNTplus reagent set (0.015 2.0 mg/L NH₃-N)
- TNT831 LR Ammonia TNTplus reagent set (1.0 12.0 mg/L NH₃-N)
- TNT832 HR Ammonia TNTplus reagent set (2.0 47.0 mg/L NH₃-N)

Chemicals:

Trapping Solution - 0.016 M H₂SO₄: 4 mLs of a 1 M H2SO4 diluted to 250 mLs with deionized water Releasing Solution - Sodium Borate (Na₂B₄O₇) Buffer solution

0.025 M borate buffer (buffer for samples and standards preserved with 2 mL of concentrated H_2SO_4/L) and 0.011 M borate buffer (buffer for samples and standards not preserved with acid)

- 1) Place a 1-L container on a top-loading balance
- 2) Zero the balance
- 3) Add 964 g of deionized water
- 4) Add and fully dissolve the sodium borate (Na₂B₄O₇), refer to the table below for quantity
- 5) Add sodium hydroxide (NaOH), refer to the table below for quantity
- 6) Put the solution in a closed container



Concentration	Sodium borate	Sodium hydroxide
0.025 M borate buffer	5 g (anhydrous)	22 g (solid)
	9.5 g (decahydrate, 10 H₂O)	22 g (solid)
0.011 M borate buffer	2.17 g (anhydrous)	36 mL of 0.1 M solution
	4.2 g (decahydrate, 10 H₂O)	36 mL of 0.1 M solution

^{*}These amounts are on a per liter basis

Procedure:

Set the MICRO DIST block temperature controller to 120°C. Allow the block to warm up; it may take up to 40 min. If the DRB200 is being used for the distillation, insert the adapter sleeves into the 20-mm wells.

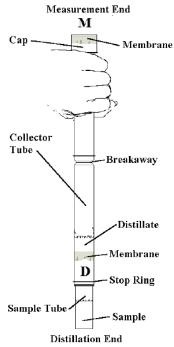


Figure 1: MICRO DIST Collector Tube

1) Using the User Filled ammonia tubes, fill the M (measuring) portion of the collector tube with 1.0 mL of $0.016 \text{ M H}_2\text{SO}_4$ for the trapping solution (Figure 1). Once the $0.016 \text{ M H}_2\text{SO}_4$ is added, take the loose membrane and press the cap over to seal in the trapping solution.



Figure 2: Placing sample in tubes



2) Place as many collector tubes as you have samples into the collector tube rack; up to 21 for one block (Micro Dist), or 8 for the DRB200. Place **6.0 mL** of sample or standard into each sample tube with a pipet (Figure 2). In this method, the standards are distilled with the samples.



Figure 3: Adding the releasing buffer solution

3) Buffer the sample or standard to pH 9.5 to decrease the hydrolysis of cyanates and organic nitrogen compounds and to convert the ammonium ion to ammonia gas during the distillation. Add 1.0 mL of 0.025 M borate buffer for samples preserved with H_2SO_4 **OR** 0.75 mL of 0.011 M borate buffer for non-preserved samples to each sample tube.

Figure 4: Starting the seal

- 4) **Immediately** push the **D** end of the collector tube over the open end of each sample tube to start the seal (Figure 4).
- 5) Place the assembly in the press (Figure 4), putting the sample tube through the hole in the white base. Before pressing, the user should grip the collector tube firmly at the breakaway point to keep the tube from shifting during the pressing procedure.
- 6) The pressing motion should be smooth, constant pressure, which is just enough to slide the sample tube inside the collector tube. A jerky, forced motion may cause added strain to the tube and could potentially crack it. Press down on the handle until the stop ring on the sample tube hits the **D** end of the collector tube.
- 7) Put on the heat-resistant gloves. Push the sample tube and **D** end of each tube all the way into the preheated block so that the collector tube stop ring touches the block.
- 8) Set the timer for **30** minutes.





Figure 5: Removing the Sample tube

- 9) When **30** min are up, put on the heat-resistant gloves. Remove the first tube from the block and **immediately** pull off its sample tube using a downward, twisting motion as opposed to a sideways motion (Figure 5). You must pull off the sample tube **within 4 s** of removing it from the block or suckback of the sample will occur (*if you are using the DRB200, see the troubleshooting section at the end of the note for instructions on the DRB200's timer*). Drop the sample tube and the hot solution left in it into a waste bucket reserved for this purpose.
- 10) Invert each collector tube and place it into the collector tube rack, now with the **D** ends up.
- 11) Allow tubes to cool for at least 10 min.

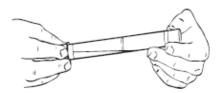


Figure 6: Rinsing the Collector tube walls

12) For each collector tube, hold the tube horizontally and rinse its walls with the distillate in order to homogenize it. Slowly roll the distillate around in the tube to gather all droplets clinging to the tube walls into the bulk of the distillate (Figure 6). Then, slowly return the collector tube to an upright position so that the **D** end is up. Stubborn drops will often fall into the **M** end when the tube is flicked with your finger or shaken down like a thermometer in a whipping motion.

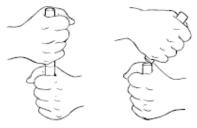


Figure 7: Breaking the 'D' End of the Collector tube

13) With the **D** end still up, break the collector tube in half by pulling the **D** end hard towards yourself to break it, then twisting and tearing off the **D** end (Figure 7). Discard the **D** end.



14) In the remaining \mathbf{M} end of the collector tube, dilute to the 6.0 mL mark with reagent water. This results in the original sample volume in 0.003 M H_2SO_4 .



Figure 8: Shaking the tube

- 15) Shake the tube with a gentle whipping motion to mix in diluent water (Figure 8). Do not invert the sample. With the **M** end down; place the tube into the collector rack. **Seal both ends of the tube with Parafilm if you will not determine the sample immediately**.
- 16) Determine the ammonia concentration by following TNT830, 831 or 832 procedure depending on the estimated concentration range. Refer to the working instructions in method 10205, TNT830-DOC316.53.01081, TNT831-DOC316.53.01082 and TNT832-DOC316.53.01083.

Troubleshooting – DRB200 Timer and Membrane Suck-back Issues

The DRB200 has a timer built into the block. Once the timer expires on the block, the temperature will start to decrease. The decreasing temperature of the block can be problematic if the distillation tubes are not removed immediately after the distillation time expires. Once the block temperature starts to decrease, a pressure difference will develop between the sample and the distillate causing the distillate to be sucked-back into the sample tube, ruining the distillate sample. To keep the digestion block from cooling off after the distillation has finished, do not start the timer on the DRB200 and track the 30-min distillation time period off-line.

Troubleshooting – Membrane Caking

When solid samples or sludges are distilled, foam comes up through the membrane, or scum cakes over the underside of the membrane causing it to be pushed up. This occurs when the sample has a lot of organics in it such as grease or oils. The scum or foam is organic surfactants which wet the hydrophobic membrane. This causes it to lose its hydrophobicity and thus not function properly. The placement of the membranes on all collector tubes is elevated such that the matrix foam normally will not come into contact with the membrane. Be careful of organic material caking the membrane or actually coming through the membrane as this will cause pressure to build up in the sample tube. The pressure is not large but it is sufficient to cause spattering of the hot sample when the sample tube is removed. In some cases the distillation membrane may pop out of the ring.



Running Solid Samples with Micro Dist

The Micro Dist is capable of handling many different kinds of solid samples from sands to sludges. As a general guideline, if the sample is high in organic content use only 0.5 g or less of sample. If the sample is low in organic content, use up to 1 g of sample. Experiment with samples to determine the best weight of sample to add for each matrix type. The sample will be diluted with DI water (5 to 6 mL) per the MicroDist manual. Calculating the amount of sample in mg/kg after analysis:

 $A = C \times ((1 L / I000 mL) \times V)$

 $B = A \text{ divided by } (W \times 1000 \text{ g/kg})$

Where:

A = mg of analyte in the sample

B = mg of analyte per kg of sample (mg/kg)

C = determined concentration (mg/L)

V = dilution volume in tube (mL)

W = weight of the original sample (g)

If foaming or caking of the membrane continues to be a problem even with reduced sample weights of 0.5 g, try the following:

- Add activated charcoal so it covers the surface of the solid and then fill the remaining void space with glass wool. When trying this procedure it would be recommended that 4-5 mL of water be used versus 6 mL.
- Test a known standard with one of these procedures and a spiked sample of the foaming or caking matrix to conclude whether these solutions will work.

Conclusion

For more information on the MicroDist block, the distillation tubes and troubleshooting, see the Lachat MicroDist User Manual, DOC022.97.80305. For more information on interferences, please see method 10205, TNT830-DOC316.53.01081, TNT831-DOC316.53.01082 and TNT832-DOC316.53.01083.

FOR TECHNICAL ASSISTANCE, PRICE INFORMATION AND ORDERING:
Tel: 800-227-4224 | E-Mail: techhelp@hach.com

To locate the HACH office or distributor serving you, visit: www.hach.com

LIT2222 Rev1

© Hach Company, 2015. All rights reserved.

In the interest of improving and updating its equipment, Hach Company reserves the right to alter specifications to equipment at any time.

